























cyclohexanone derivative **1** with *trans*-substitution at C(3) and C(4) leads to the *cis*-octahydrophenanthrene **2** [3] (Scheme 1). Instigated by this result based on an X-ray diffraction analysis, the closely related 5,8-dimethoxy derivative **3** was analyzed by ¹H-NMR

Table. Structures and ¹H-NMR Data (300 MHz) of 1,2,3,4,4a,9,10,10a-Octahydrophenanthrenes²⁾

Compound	No.	Ref.	$J(4a,10a)$ [Hz]/ $\delta(H-C(10a))$ [ppm]	Compound	No.	Ref.	$J(4a,10a)$ [Hz]/ $\delta(H-C(10a))$ [ppm]
	5	[2] [6]	5.0/2.96		6		11.5/2.35
	7 ^{a)}	[7]	5.5/3.92		8 ^{b)}	[7]	12.0/3.28
R, R = Phthaloyl							
	9	[2] [7]	4.5/2.68		17	[12]	7.0/3.21 ^{c)}
	10 ^{d)}	[7]	5.0/3.02		18	[2] [9] [10]	–/ ^{e)}
	11	[8]	5.0/2.92		19 ^{f)}	[2]	11.0/ca. 1.5
	12	[8]	4.5/2.66		20 ^{g)}	[11]	11.0/–
	13	[2]	5.0/2.90		21 ^{h)}	[2]	11.5/2.08
	14	[2]	5.0/ca. 2.45		22	[2] [6] [13]	12.0/– ⁱ⁾
	15	[2]	4.5/2.53		1	[3]	11.5/– ^{k)}
	3	[4]	5.5/2.92		2	[3]	5.5/2.94
R = CH ₃				R = CH ₃ , R ¹ = CH ₂ CO ₂ CH ₃			
	4	[4]	5.3/2.99		16	[4]	11.5/ca. 2.2

^{a)} $J(10, 10a) = 13.5$ Hz. ^{b)} $J(10,10a) = 12.5$ Hz. ^{c)} $J(3a,9a)/\delta(H-C(9a))$. ^{d)} $J(10,10a) = 13.5$ Hz. ^{e)} $J(4,4a)$ could not be determined due to the overlapping signals of H–C(4a) and H_{eq}–C(4); its *trans*-configuration has, however, been proven unambiguously before [2] [9] [10]. ^{f)} The assignment of the H_{eq}–C(4) and H–C(4a) signals has been achieved by ¹H, ¹³C decoupling experiments. ^{g)} $J(4,4a) = < 5$ Hz. ^{h)} $J(4,4a) = 10$ Hz. ⁱ⁾ $J(1',2')$. ^{k)} $J(3,4)$.

²⁾ For a detailed spectral description, cf. *Exper. Part*.

[3a]. In contrast to previous reports, in which the *trans*-configuration was assigned, wholesale but through an erroneous generalization, to all 1,2,3,4,4a,9,10,10a-octahydrophenanthrenes, except the 5,8-dimethyl-4,9-dione **4** [4] (*Scheme 1*), the 5,8-dimethoxy compound **3** was found to have the *cis*-configuration as well [3a]. An inspection of the premises, which apparently led to erroneous assignments in the *Elad-Ginsburg* morphine synthesis [5], was necessary. This synthesis [5] was successful, and it eventually led to products of correct configuration. Nevertheless, it should presumably have been based upon correct configurational assignments and transformations of such octahydrophenanthrene systems. To answer the questions raised, we decided to determine the relative configuration of these intermediates and model compounds ([2–13]) by NMR analysis. The structures **1–22** with their appropriate relative configurations, based upon the magnitude of the coupling constant between H–C(4a) and H–C(10a), $J(4a,10a) = 4.5–5.5$ Hz for *cis*-, $J(4a,10a) = 11–12$ Hz for *trans*-anelated isomers, are listed in the *Table*².

Surprisingly, not only compounds with C(5)-substitution as well as a C(4)-oxo function (see **2**, **3**, **4**, **13**, **14**) and respective acetal derivatives (**15**) had the *cis*-configuration; this list also includes derivatives without C(5)-substituent (see **5**, **7**, **9**, **10**, **11**, **12**), which turned out to be *cis* or epimer mixtures with the *cis*-isomer as the major component. With a $J(3a,9a)$ value of 7 Hz, the 1,2,3,3a,8,9,9a-hexahydroindene-3,8-dione (**17**) [12] is the *cis*-isomer as well. The *trans*-configuration of the 2'-aryl-cyclohexylacetic-acid precursors was found to be retained in octahydrophenanthrenes without oxo function at C(4) (see **18**, **19**, **20**, **21**) (*Table*). Since the 2'-cyclohexylacetic acids were obtained from intermediates containing the corresponding carbonyl group, it is reassuring to know that *Barton's* conformational tenets, regarding stability of the *trans*-1,2-diequatorial isomer, hold here as well.

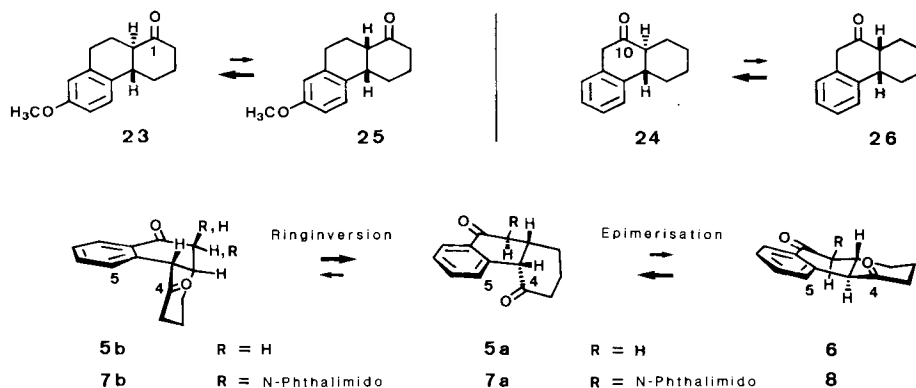
To obtain an estimate of the energy difference between the tricyclic *cis*- and the *trans*-system, equilibration of the parent 1,2,3,4,4a,9,10,10a-octahydrophenanthrene-4,9-dione (obtained as a 82.5:17.5 mixture of *cis*- and *trans*-isomers **5** and **6** by hydrolysis of acetal **9**) was studied. The pure isomers could be obtained and characterized by HPLC separation of the mixture. According to the physical data, either the *cis*-isomer **5** or an equilibrated mixture of **5** and **6** correspond to the material described previously [2] [6]³. Heating solutions of either pure **5** or pure **6** in toluene/EtOH containing a catalytic amount of CH₃SO₃H led to the same mixture of 82% *cis*-isomer **5** and 18% *trans*-isomer **6**⁴. This ratio, therefore, reflects the thermodynamic equilibrium of **5** and **6**. The 10-phthalimidooctahydrophenanthrene-4,9-dione, obtained by two independent methods [7], turned out to be an 8:2 mixture of *cis*-isomer **7** and *trans*-epimer **8** as well. In this case, however, no separation was attempted.

It is interesting to note, that in the case of 1,2,3,4,4a,9,10,10a-octahydrophenanthrenes with a C(1)- or a C(10)-oxo function the *trans*-isomers **23** and **24** are energetically favored as compared to the *cis*-isomers **25** and **26** (**23/25** = 4:1 [14]; **24/26** = 61:39 [15],

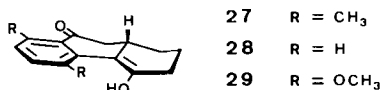
³) Recrystallization from CH₂Cl₂/Et₂O/hexane gives a melting point of 80–82° for the *cis*-isomer **5** and 109–112° for the *trans*-isomer **6**. In the case of **5**, redetermination of the melting point, after the material had resolidified, gave a higher reading (94–97°) which is close to the reported value (94–95°, either from BuOH [2] or from dilute AcOH [6]). It was not verified, whether this higher melting point is due to a different crystal modification or to *cis/trans*-isomerization.

⁴) The ratio was determined from the ¹H-NMR spectra by integration of the H–C(4a) signals.

Scheme 2

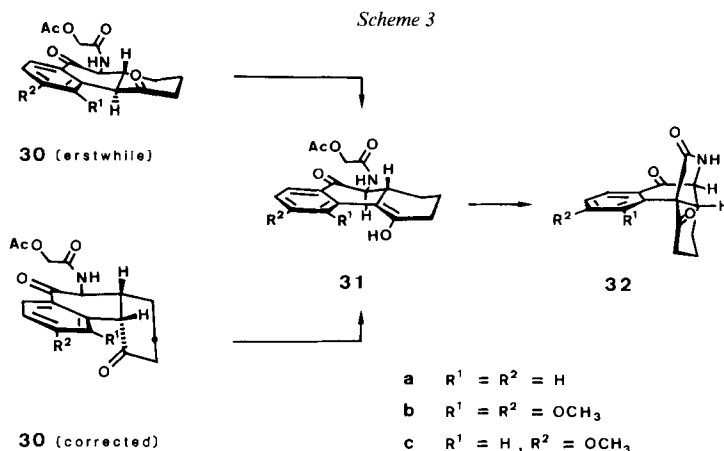


(Scheme 2). Probably, the preference of the *cis*-epimers **5** and **7** over the *trans*-isomers **6** and **8** is due to the oxo group at C(4), whose steric interaction with C(5) is less severe for the conformers **5a/7a** than for the conformers **5b/7b** or the *trans*-isomers **6/8** (Scheme 2). This strain increases for C(5)-substituted derivatives, and in these cases (see **2**, **3**, **4**, **13**, **14**, Table), the *trans*-isomers cannot be detected in equilibrated mixtures. In general, the acidic conditions applied for the cyclization of *trans*-substituted 2'-aryl-3'-oxocyclohexylacetic acids (e.g. **1**, **22**, Table) suffice for complete equilibration⁵). A notable exception is, however, 2'-(2'',5''-dimethylphenyl)-3'-oxocyclohexylacetic acid, which affords a mixture of *trans*- and *cis*-octahydrophenanthrenes **16** and **4**, respectively, with the thermodynamically less stable *trans*-isomer **16** predominating upon cyclization with liquid HF [4]. The kinetic stability of **16** must, therefore, be due to the steric strain of the enol intermediate **27**, which is higher in energy than the unsubstituted or MeO-substituted derivatives **28** and **29**.



As shown before by X-ray and ¹H-NMR analysis [3a], *cis*-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-4,9-diones prefer the chair/chair conformation represented by the formulae **5a/7a** (Scheme 2). This is fully confirmed by the NMR spectra of the C(10)-substituted compounds **7** and **10** of this study. The *J*(10,10a) value of 13.5 Hz corresponds to an *anti-periplanar* (trans-diaxial) relation, which is only possible for the conformer **7a** with an equatorial *exo*-substituent R (Scheme 2). The δ (H-C(10a)) value listed in the Table is another characteristic feature of these ¹H-NMR data, which is of analytical importance for those octahydrophenanthrenes, whose configuration cannot be determined by *J*(4a,10a). By comparing the epimeric pairs **5/6**, **7/8**, **4/16**, it can be seen that δ (H-C(10a)) is shifted by ca. 0.6 ppm to lower field for the *cis*-isomers **5**, **7**, and **4**. Further, δ (H-C(10a)) is influenced by substituents at C(10) (compare **5** and **7**, **9** and **10**, **6** and **8**) and by the C(9)-oxo group (compare **5** and **9**, **11** and **12**, **13** and **15**).

⁵) It is here that Ginsburg and Pappo [2] made a wrong assumption continued for the MeO-substituted analogs [5].



The key-sequence of the *Elad-Ginsburg* synthesis of morphine [5] is depicted in *Scheme 3* according to [5], but we now must correct the structure of **30** as a result of the data presented herein. Cyclization of **30a** in model studies for the morphine synthesis were interpreted to proceed *via* the enol **31a** (*cf.* [2] [8]); it was no surprise, that **32a** was correlated with *N*-methylmorphinan (of *cis*-configuration) rather than with *N*-methylisomorphinan, the *trans*-isomer thereof. In fact, the then believed *trans*-configuration in the intermediate octahydrophenanthrenes [2] [7] never caused any worry, since eventually the tetracyclic systems constructed (*e.g.* **32a, b** [5]) were expected to have the *cis*-configuration (after epimerization *via* **31a, b**), which *Nature* has produced in morphine and codeine. *Stork* has proposed an ingenious mechanism for the unprecedented cyclization **30** to **32** *cum* epimerization through **31** [16]. The crucial point for the correct stereochemical outcome of this synthesis is, however, the relative configuration at C(10) and C(10a) of **30**. The rationalization given by *Stork* [16] for a *trans*-configured intermediate – ‘the amino group at C(10) must be equatorial, since its adjacency to a ketone allows it to epimerize to the more stable configuration after its formation’ – is valid for the *cis*-configured octahydrophenanthrenes as well, only because this system prefers the ψ -chair-chair conformation **5a** depicted in *Scheme 2* and not the ring-inverted ψ -chair-chair conformation **5b** (*cf.* [3a]).

Finally, we must issue a warning. Unfortunately, we cannot include *all* of the intermediates in the *Elad-Ginsburg* morphine synthesis [5] and similar work cited herein. We measured the ¹H-NMR spectra of a random sampling of intermediates, but we cannot guarantee that *not one* of the tricyclic compounds involved there does not retain *trans*-configuration. If any of these intermediates is prepared again, there are now available two formidable tools, X-ray structure analysis and ¹H-NMR spectroscopy, to check correct configuration.

Postscript: No responsibility for whatever errors exist in [2] [5] [7] is to be attributed to the late *D. Elad* or to *R. Pappo*. It is solely that of one of us (*D.G.*).

This work was supported by *Ciba-Geigy AG*, Basel. We are indebted to the following persons of the analytical department of the *ETH Zürich*: Ms. *B. Brandenburg* and Mr. *F. Fehr* (NMR), Prof. *J. Seibl* and Mrs. *L. Golgowsky* (MS).

Experimental Part

1. Separation and Isomerization of *cis*- and *trans*-1,2,3,4,4a,9,10,10a-Octahydrophenanthrene-4,9-dione (5 and 6, respectively). – a) *Separation*. A mixture 5/6 (100 mg, 5/6 = 82.5:17.5) was separated by HPCL (silica gel, hexane/CH₂Cl₂/Et₂O 5:5:1, 25 bar, 20 ml/min flow rate, UV detection at 250 nm) giving 10 mg of (4aR*,10aS*)-epimer 6 (*trans*, Vol._{ret.} = 356 ml, m.p. 109–112° from CH₂Cl₂/Et₂O/hexane) and 69 mg of (4aS*,10aS*)-epimer 5 (*cis*, Vol._{ret.} = 428 ml, m.p. 80–82° from CH₂Cl₂/Et₂O/hexane, remelting at 94–97° after resolidification).

b) *Isomerization of 6 (trans)*. A soln. of 6 (ca. 3.5 mg) in 0.4 ml of toluene and 25 µl of 2% CH₃SO₃H in EtOH was stirred under reflux for 1½ h at 100°. Workup with Et₂O and chromatography (silica gel, hexane/CH₂Cl₂/Et₂O 9:9:2) gave 2.7 mg of 5/6 (82.2:17.8 according to ¹H-NMR, integration of H–C(4a)).

c) *Isomerization of 5 (cis)*. A soln. of 5 (ca. 8 mg) in 1.5 ml of toluene and 75 µl of 2% CH₃SO₃H in EtOH was stirred under reflux at 100° for 1½ h. Workup and purification as above gave 6 mg of 5/6 (82:18 according to ¹H-NMR, integration of H–C(4a)).

2. NMR Spectra of 1–22 (cf. the Table). – The following spectra have been measured on a Bruker WM-300 spectrometer at 300 MHz (¹H) and 75.4 MHz (¹³C).

Dimethyl [(1R,3S*,4R*)-4-(5'-Acetoxy-2',4'-dimethoxyphenyl)-5-oxocyclohexane-1,3-diacetate (1) [3].* ¹H-NMR (300 MHz, CDCl₃): 1.82 (ddd, *J* = 14, 12, 4.5), 1.98 (*d*, *J* = 14, further split, *w*_{1/2} ≈ 8) (2 H–C(2)); 2.14 (*dd*, *J* = 16, 8.5), 2.30 (*dd*, *J* = 16, 4), 2.46 (ddd, *J* = 15, 3.5, 2), 2.47 (*d*, *J* = 8, further split, *w*_{1/2} ≈ 2), and 2.61 (*dd*, *J* = 15, 5.5, further split, *w*_{1/2} ≈ 2) (2 H–C(6), 2 CH₂CO₂CH₃); 2.27 (*s*, CH₃CO₂–C(5')); 2.67–2.87 (*m*, H–C(1), H–C(3)); 3.43 (*d*, *J* = 11.5, further split, *w*_{1/2} ≈ 2, H–C(4)); 3.57, 3.70, 3.79, 3.83 (4s, 4 CH₃O); 6.53 (*m*, *w*_{1/2} ≈ 1.5, H–C(3')); 6.70 (*s*, H–C(6')).

Methyl [(2R,4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-8-hydroxy-5,7-dimethoxy-4,9-dioxophenanthrene-2-yl]acetate (2) [3].* ¹H-NMR (300 MHz, CDCl₃): 1.92–1.99 (*m*, 4 main signals, 2 H–C(1)); 2.35–2.63 (*m*, 7 H); 2.94 (*dt*, *J* ≈ 12, 6, 3.5, H–C(10a)); 3.71, 3.76, 3.92 (3s, 3 CH₃O); 4.15 (*d*, *J* = 5.5, H–C(4a)); 6.81 (*m*, *w*_{1/2} ≈ 2, H–C(6)); 12.02 (*d*, *J* ≈ 0.5, OH).

(4aS,10aR*)-1,2,3,4,4a,9,10,10a-Octahydro-5,8-dimethoxyphenanthrene-4,9-dione (3) [3a] [4].* ¹H-NMR (300 MHz, CDCl₃): 1.76–2.21 (*m*, 4 H); 2.37–2.65 (*m*, 2 H); 2.40 (ddd, *J* = 16, 4.5, 1.5, H_{exo}–C(10)); 2.59 (*tdd*, *J* = 13, 6, 1, H_{ax}–C(3)); 2.92 (*d*, *J* ≈ 13.5, further split, *w*_{1/2} ≈ 12, H–C(10a)); 3.75, 3.86 (2s, 2 CH₃O); 4.27 (*d*, *J* ≈ 5.5, further split, *w*_{1/2} ≈ 3, H–C(4a)); 6.89, 7.04 (2*d*, *J* = 9, H–C(6), H–C(7)).

(4aS,4aR*)-1,2,3,4,4a,9,10,10a-Octahydro-5,8-dimethylphenanthrene-4,9-dione (4) [3a] [4].* ¹H-NMR (300 MHz, CDCl₃): 1.86 (*d*, *J* ≈ 14, further split, *w*_{1/2} ≈ 7, 1 H); 1.92–2.25 (*m*, 3 H); 2.14 (*s*, CH₃–C(5)); 2.43 (ddd, *J* = 16, 5.5, 1.5, H_{eq}–C(10)); 2.49–2.64 (*m*, 3 H); 2.95 (*s*, CH₃–C(8)); 2.99 (*d*, *J* = 14, further split, *w*_{1/2} ≈ 13, H–C(10a)); 4.06 (*d*, *J* = 5.3, further split, *w*_{1/2} ≈ 3, H–C(4a)); 7.08, 7.24 (2*d*, *J* ≈ 8, H–C(6), H–C(7)).

(4aS,10aS*)-1,2,3,4,4a,9,10,10a-Octahydrophenanthrene-4,9-dione (5) [2] [6].* ¹H-NMR (300 MHz, CDCl₃): 1.70–1.88 (1 H), 1.92–2.10 (3 H) (2*m*, 2 H–C(1), 2 H–C(2)); 2.38–2.55 (*m*, 2 H–C(3)); 2.55–2.65 (*m*, 2 H–C(10)); 2.90–3.04 (2.96) (*m*, H–C(10a)); 3.98 (*d*, *J* = 5, further split, *m*, *w*_{1/2} ≈ 2, H–C(4a)); 7.10 (*d*, *J* ≈ 7.5, further split, *m*, *w*_{1/2} ≈ 3, H–C(5)); 7.40 (*t*, *J* ≈ 7.5, further split, *m*, *w*_{1/2} ≈ 3), 7.54 (*td*, *J* ≈ 7.5, 1.5) (H–C(6), H–C(7)); 8.08 (*dd*, *J* ≈ 7.5, 1.5, H–C(8)).

(4aR,10aS*)-1,2,3,4,4a,9,10,10a-Octahydrophenanthrene-4,9-dione (6).* ¹H-NMR (300 MHz, CDCl₃): 1.75–2.05 (3 H), 2.15–ca. 2.3 (1 H) (2*m*, 2 H–C(1), 2 H–C(2)); ca. 2.25–2.45 (*m*, H–C(10a)); 2.60 (*dd*, *J* = 16, 13), 2.79 (*dd*, *J* = 16, 3.5) (2 H–C(10)); 2.53–2.71 (*m*, 2 H–C(3)); 3.89 (*d*, *J* = 11.5, further split, *m*, *w*_{1/2} ≈ 3, H–C(4a)); 7.38 (*t*, *J* ≈ 7.5, further split, *m*, *w*_{1/2} ≈ 4, H–C(7)); 7.5–7.65 (*m*, H–C(5), H–C(6)); 8.05 (*d*, *J* ≈ 7.5, further split, *m*, *w*_{1/2} ≈ 3, H–C(8)).

N-[(4aS,10S*,10aR*)- and N-[(4aR*,10S*,10aR*)-1,2,3,4,4a,9,10,10a-Octahydro-4,9-dioxophenanthrene-10-yl]phthalimide (7 and 8, respectively) [7].* ¹H-NMR (300 MHz, CDCl₃): 1.75–2.3 (*m*, 2 H–C(1'), 2 H–C(2')); 2.5–2.75 (*m*, 2 H–C(3')); 3.2–3.36 (3.28) (*m*, H–C(10a')), (4a'R*)-epimer); 3.86–3.99 (3.92) (*m*, H–C(10a')), (4a'S*)-epimer); ca. 4.12 (*d*, *J* ≈ 12, H–C(4a')), (4a'R*)-epimer); 4.15 (*d*, *J* = 5.5, H–C(4a')), (4a'S*)-epimer); 4.94 (*d*, *J* = 13.5, H–C(10')), (4a'S*)-epimer); 5.05 (*d*, *J* = 12.5, H–C(10')), (4a'R*)-epimer); 7.1–7.25, 7.35–7.5, 7.5–7.7 (3*m*, H–C(5'), H–C(6'), H–C(7')); 7.7–7.8 (2 H), 7.8–7.95 (2 H) (2*m*, H–C(3), H–C(4), H–C(5), H–C(6)); 8.05–8.15 (*m*, H–C(8')).

(4aS,10aS*)-1,2,3,4,4a,9,10,10a-Octahydrophenanthrene-4-spiro-2'-(1',3'-dioxolane)-9-one (9) [2] [7].* ¹H-NMR (300 MHz, CDCl₃): 1.56–1.95 (*m*, 2 H–C(1), 2 H–C(2), 2 H–C(3)); 2.35 (ddd, *J* = 18, 5.5, 1), 3.19 (*dd*, *J* = 18, 14) (2 H–C(10)); 2.61–2.75 (2.68) (*m*, H–C(10a)); 2.90–3.02 (1 H), 3.16–3.27 (1 H), 3.57–3.72 (2 H) (3*m*, 2 H–C(4'), 2 H–C(5')); 3.15 (*d*, *J* ≈ 4.5, H–C(4a)); 7.27–7.38 (1 H), 7.40–7.48 (2 H) (2*m*, H–C(5), H–C(6), H–C(7)); 8.04 (*d*, *J* ≈ 7.5, further split, *m*, *w*_{1/2} ≈ 3, H–C(8)).

Ethyl [(4aS,10S*,10aR*)-1,2,3,4,4a,9,10,10a-Octahydro-9-oxophenanthrene-4-spiro-2'-(1',3'-dioxolane)-10-yl]carboxylate (10)* [7]. ¹H-NMR (300 MHz, CDCl₃): 1.31 (t, *J* = 7, CH₃CH₂O); 1.57–1.78 (4 H), 1.78–1.98 (2 H) (2m, 2 H–C(1'), 2 H–C(2'), 2 H–C(3')); 2.92–3.02 (1 H), 3.14–3.23 (1 H), 3.58–3.74 (2 H) (3m, 2 H–C(4''), 2 H–C(5'')); 2.95–3.08 (3.02) (m, H–C(10a'')); 3.22 (d, *J* = 5, H–C(4a'')); 4.22, 4.28 (2dq, *J* = 11, 7, CH₃CH₂O); 4.30 (d, *J* = 13.5, H–C(10'')); 7.32–7.40 (m, 3 main signals, H–C(7'')); 7.40–7.53 (m, H–C(5'), H–C(6'')); 8.04 (d, *J* = 8, further split, m, *w*_{1/2} ≈ 3, H–C(8')).

(4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-6-methoxyphenanthrene-4,9-dione (11) [8]. ¹H-NMR (300 MHz, CDCl₃): 1.7–1.85 (1 H), 1.88–2.1 (3 H) (2 H–C(1), 2 H–C(2)); 2.4–2.5 (m, 2 H–C(3)); 2.5–2.63 (m, 2 H–C(10)); 2.85–2.98 (m, H–C(10a)); 3.83 (s, CH₃O); 3.94 (d, *J* = 5, H–C(4a)); 6.54 (d, *J* ≈ 2.5, further split by a small coupling, H–C(5)); 6.90 (dd, *J* = 8.5, 2.5, further split by a small coupling, H–C(7)); 8.05 (d, *J* = 8.5, H–C(8)).

(4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-6-methoxyphenanthrene-4-spiro-2'-(1',3'-dioxolane)-9-one (12) [8]. ¹H-NMR (300 MHz, CDCl₃): 1.55–1.94 (m, 2 H–C(1), 2 H–C(2), 2 H–C(3)); 2.30 (ddd, *J* = 18, 5.5, ca. 1), 3.15 (dd, *J* = 18, 14) (2 H–C(10)); 2.59–2.72 (m, H–C(10a)); 2.98–3.07 (1 H), 3.3–3.4 (1 H), 3.6–3.76 (2 H) (2 H–C(4'), 2 H–C(5')); 3.1 (d, *J* = 4.5, H–C(4a)); 3.85 (s, CH₃O); 6.86 (dd, *J* = 8.5, 2.5, H–C(7)); 6.94 (d, *J* = 2.5, H–C(5)); 8.01 (d, *J* = 8.5, H–C(8)).

(4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-5,6-dimethoxyphenanthrene-4,9-dione (13) [2]. ¹H-NMR (300 MHz, CDCl₃): 1.75–1.87 (m, H_{eq}–C(1)); 1.9–2.2 (m, 2 H–C(2), H_{ax}–C(1)); 2.37–2.65 (m, 2 H–C(3), 2 H–C(10)); 2.84–2.97 (m, H–C(10a)); 3.76, 3.93 (2s, 2 CH₃O); 4.26 (d, *J* = 5, H–C(4a)); 6.96 (d, *J* = 8.5, H–C(7)); 7.88 (d, *J* = 8.5, H–C(8)).

(4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-5,6-dimethoxyphenanthrene-4-one (14) [2]. ¹H-NMR (300 MHz, CDCl₃): 1.56–1.68 (2 H), 1.74–1.87 (1 H), 1.87–2.14 (3 H) (3m, 2 H–C(1), 2 H–C(2), 2 H–C(10)); 2.35–2.58 (m, 2 H–C(9), H–C(10a)); 3.73, 3.83 (2s, 2 CH₃O); 4.00 (d, *J* = 5, further split, *w*_{1/2} ≈ 2, H–C(4a)); 6.80 (s, H–C(7), H–C(8)).

(4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-5,6-dimethoxyphenanthrene-4-spiro-2'-(1',3'-dioxolane)-9-one (15) [2]. ¹H-NMR (300 MHz, CDCl₃): 1.55–2.0 (m, 2 H–C(1), 2 H–C(2), 2 H–C(3)); 2.28 (ddd, *J* = 18, 7, ca. 1), 3.04 (dd, *J* = 18, 13) (2 H–C(10)); 2.47–2.60 (m, H–C(10a)); 2.97–3.05 (1 H), 3.18–3.27 (1 H), 3.6–3.72 (2 H) (2 H–C(4'), 2 H–C(5')); 3.66 (d, *J* = 4.5, H–C(4a)); 3.90, 3.92 (2s, 2 CH₃O); 6.92 (d, *J* = 8.5, H–C(7)); 7.78 (d, *J* = 8.5, H–C(8)).

(4aR*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-5,8-dimethylphenanthrene-4,9-dione (16) [4]. ¹H-NMR (300 MHz, CDCl₃): 1.79–2.07 (m, 3 H); 2.04 (s, CH₃–C(5)); 2.14–2.33 (m, 2 H); 2.49–2.79 (m, 4 H); 2.57 (s, CH₃–C(8)); 3.99 (d, *J* = 11.5, further split, *w*_{1/2} ≈ 2.5, H–C(4a)); 7.09, 7.26 (2d, *J* ≈ 8, H–C(6), H–C(7)).

(3aS*,9aS*)-1,3,3a,8,9,9a-Hexahydrobenz[e]indeno-3,8-dione (17) [12]. ¹H-NMR (300 MHz, CDCl₃): 1.85–1.98, 2.17–2.32 (2m, 2 H–C(1)); 2.33–2.58 (m, 2 H–C(2)); 2.55 (dd, *J* = 16, 10.5), 2.81 (dd, *J* = 16, 5.5, further split, m, *w*_{1/2} ≈ 2) (2 H–C(9)); 3.14–3.28 (3.21) (m, H–C(9a)); 3.58 (d, *J* ≈ 7, further split, m, *w*_{1/2} ≈ 3, H–C(3a)); 7.32–7.45, 7.54–7.64 (2m, 3 main peaks each, H–C(5), H–C(6)); 7.47–7.54 (m, 2 main peaks, H–C(4)); 7.99 (dd, *J* ≈ 8, 1, H–C(7)).

(4aR*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydrophenanthrene-9-one (18) [2] [9] [10]. ¹H-NMR (300 MHz, CDCl₃): 1.20–1.61 (4 H), 1.72–1.93 (3 H), 1.93–2.04 (1 H) (3m, 2 H–C(1), 2 H–C(2), 2 H–C(3), H_{ax}–C(4), H–C(10a)); 2.37 (dd, *J* = 17, 13), 2.68 (dd, *J* = 17, 3.5) (2 H–C(10)); 2.49–2.62 (m, H_{eq}–C(4), H–C(4a)); 7.32 (t, *J* = 7.5, further split, *w*_{1/2} ≈ 3, H–C(7)); 7.43 (d, *J* ≈ 8, further split, *w*_{1/2} ≈ 3, H–C(5)); 7.52 (t, *J* = 7.5, further split, *w*_{1/2} ≈ 3, H–C(6)); 8.06 (dd, *J* = 8, 1.5, H–C(8)).

(4aR*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-5,6-dimethoxyphenanthrene-9-one (19) [2]. ¹H-NMR (300 MHz, CDCl₃): 1.05–1.23 (m, H_{ax}–C(4)); 1.3–1.65 (3 H), 1.75–2.0 (4 H) (2 H–C(1), 2 H–C(2), 2 H–C(3), H–C(10a)); 2.33 (dd, *J* = 15, 13.5), 2.45 (dd, *J* = 15, 3) (2 H–C(10)); 2.71 (td, *J* = 11, 3, H–C(4a)); 3.19 (dq, *J* = 13, ca. 3, H_{eq}–C(4)); 3.79, 3.91 (2s, 2 CH₃O); 6.89 (d, *J* = 8.5, H–C(7)); 7.86 (d, *J* = 8.5, H–C(8)). ¹³C-NMR (75 MHz, CDCl₃): 26.2, 27.3 (C(2), C(3)); 30.3 (C(4): t, off-res. ¹H-dec., d, upon selective ¹H-decoupling at 957 Hz/3.19 ppm); 33.8 (C(1)); 40.8 (C(10a)); 43.7 (C(4a): d, off-res. ¹H-dec., s, upon selective ¹H-decoupling at 813 Hz/2.71 ppm); 45.4 (C(10)); 55.7, 59.9 (2 CH₃O); 110.1 (C(7)); 124.1 (C(8)); 127.3 (C(5a)); 139.4 (C(8a)); 147.6 (C(5)); 157.4 (C(6)); 196.8 (C(9)).

(4R*,4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydrophenanthrene-4-yl Acetate (20) [11]. ¹H-NMR (300 MHz, CDCl₃): 1.1–1.3 (1 H), 1.4–1.75 (4 H), 1.77–1.96 (3 H), 2.05–2.2 (1 H, not H–C(10a), presumably H–C(3)) (4m, 2 H–C(1), 2 H–C(2), 2 H–C(3), 2 H–C(10), H–C(10a)); 1.86 (s, CH₃COO–C(4)); 2.55 (d, *J* ≈ 11, further split, m, *w*_{1/2} ≈ 5, H–C(4a)); 2.75–3.0 (m, 2 H–C(9)); 5.82 (m, *w*_{1/2} ≈ 7, H–C(4)); 7.02–7.15 (3 H), 7.15–7.25 (1 H) (2m, H–C(5), H–C(6), H–C(7), H–C(8)).

(4S*,4aS*,10aS*)-1,2,3,4,4a,9,10,10a-Octahydro-5,6-dimethoxy-9-oxophenanthrene-4-yl Acetate (21) [2]. ¹H-NMR (300 MHz, CDCl₃): 1.16–1.54 (3 H), 1.73–1.95 (2 H), 2.32–2.43 (1 H) (3m, 2 H–C(1'), 2 H–C(2)'),

H–C(3'')); 2.02 (*s*, CH₃COO–C(4'')); 2.08 (*qt*, $J \approx 12$, 4.5, H–C(10a'')); 2.32 (*dd*, $J = 18$, 12), 2.76 (*dd*, $J = 18$, 4.5) (2 H–C(10'')); 2.81 (*dd*, $J \approx 11.5$, 10, H–C(4a'')); 3.71, 3.91 (2*s*, 2 CH₃O); 5.28 (*td*, $J \approx 10$, 4.5, H–C(4'')); 6.88 (*d*, $J = 8.5$, H–C(7'')); 7.81 (*d*, $J = 8.5$, H–C(8'')).

[(1*S**,2*R**)-3-Oxo-2-phenylcyclohexyl]acetic Acid (**22**) [2] [6] [13]. ¹H-NMR (300 MHz, CDCl₃): 1.58–1.75 (1 H), 1.75–1.94 (1 H), 2.08–2.24 (2 H) (3*m*, 2 H–C(5'), 2 H–C(6'')); 2.10 (*dd*, $J = 16$, 9), 2.23 (*dd*, $J = 16$, 4) (2 H–C(2)); 2.37–2.63 (*m*, 2 H–C(4'), H–C(1'')); 3.40 (*d*, $J = 12$, H–C(2'')); 7.03–7.12 (2 H), 7.20–7.38 (3 H) (2*m*, Ph); 7.2–8.7 (br., COOH).

REFERENCES

- [1] A. J. Floyd, S. F. Dyke, S. E. Ward, *Chem. Rev.* **1976**, *76*, 509.
 [2] D. Ginsburg, R. Pappo, *J. Chem. Soc.* **1951**, 938.
 [3] a) R. O. Duthaler, P. Mathies, W. Petter, Ch. Heuberger, V. Scherrer, *Helv. Chim. Acta* **1984**, *67*, 1217; b) R. O. Duthaler, Ch. Heuberger, U. H.-U. Wegmann, V. Scherrer, *Chimia* **1985**, *39*, 174.
 [4] Sh. Bien, L. Cohen, K. Scheinmann, *J. Chem. Soc.* **1965**, 1495.
 [5] a) D. Elad, D. Ginsburg, *J. Am. Chem. Soc.* **1954**, *76*, 312; b) *J. Chem. Soc.* **1954**, 3052.
 [6] C. F. Koelsch, *J. Am. Chem. Soc.* **1951**, *73*, 2951.
 [7] D. Ginsburg, R. Pappo, *J. Chem. Soc.* **1953**, 1524.
 [8] Sh. Bien, D. Ginsburg, *J. Chem. Soc.* **1963**, 2065.
 [9] D. Gutsche, W. S. Johnson, *J. Am. Chem. Soc.* **1946**, *68*, 2239.
 [10] E. Buchta, H. Ziener, *Liebigs Ann. Chem.* **1956**, *601*, 155.
 [11] R. Pappo, D. Ginsburg, unpublished.
 [12] Y. Amiel, A. Löffler, D. Ginsburg, *J. Am. Chem. Soc.* **1954**, *76*, 3625.
 [13] W. E. Bachmann, E. J. Fornefeld, *J. Am. Chem. Soc.* **1950**, *72*, 5529.
 [14] a) A. J. Birch, H. Smith, R. E. Thornton, *J. Chem. Soc.* **1957**, 1339; b) D. Varech, L. Lacombe, J. Jacques, *Nouv. J. Chim.* **1984**, *8*, 445.
 [15] a) J. T. Valko, J. Wolinsky, *J. Org. Chem.* **1979**, *44*, 1502; b) W. E. Parham, L. E. Czuba, *J. Am. Chem. Soc.* **1968**, *90*, 4030.
 [16] G. Stork, in 'The Alkaloids', Ed. R. H. F. Manske, Academic Press, New York, 1969, Vol. VI, p. 241.